

**Hybridization stripping solution**

200 mM Tris-Cl, pH 7.0  
0.1× SSC (APPENDIX 2)  
0.1% (w/v) SDS

**Nucleotide mix**

2.5 mM ATP  
2.5 mM CTP  
2.5 mM GTP  
20 mM Tris-Cl, pH 7.5  
Store at -20°C

**COMMENTARY****Background Information**

Hybridization between complementary nucleotides was implicit in the Watson-Crick model for DNA structure and was initially exploited, via renaturation kinetics, as a means of studying genome complexity. In these applications, the two hybridizing molecules were both in solution—an approach that is still utilized in “modern” techniques such as cDNA library screening, differential display, and cDNA microarray (UNIT 6.3). The innovative idea of immobilizing one hybridizing molecule on a solid support was first proposed by Denhardt (1966) and led to methods for identification of specific sequences in genomic DNA (dot blotting; UNIT 6.4) and recombinant clones (UNIT 6.3 & 6.4). A second dimension was subsequently introduced by Southern (1975), who showed how nucleic acid molecules contained in an electrophoretic gel could be transferred to a membrane (UNIT 6.4), enabling genetic information relating to individual restriction fragments to be obtained by hybridization analysis.

Since the pioneering work of Denhardt and Southern, advances in membrane hybridization have been technical rather than conceptual. The methods have become more sophisticated, and the factors that influence hybrid stability and hybridization rate.

Hybrid stability is expressed as the melting temperature or  $T_m$ , which is the temperature at which the probe dissociates from the target DNA. For DNA-DNA hybrids, the  $T_m$  can be estimated from the equation of Meinkoth and Davidson (1984):

$$T_m = 69.3^\circ\text{C} + 16.6(\log M) + 0.41(\%GC) - 0.61(\%form) - 500/L$$

for DNA-DNA hybrids from the equation of Meinkoth and Davidson (1977):

$$T_m = 69.3^\circ\text{C} + 18.5(\log M) + 0.58(\%GC) - 11.8(\%GC)^2 - 0.56(\%form) - 820/L$$

where  $M$  is the molarity of monovalent cations, and  $L$  is the length of the hybrid in base pairs.

Since nucleotides in the DNA, %form is the percentage of formamide in the hybridization solution, and  $L$  is the length of the hybrid in base pairs. The practical considerations that arise from these two equations are summarized in Table 2.10.2A.

The second important consideration in hybridization analysis is the rate at which the hybrid is formed. Hybrid formation cannot occur until complementary regions of the two molecules become aligned, which occurs purely by chance; however, once a short nucleating region of the duplex has formed, the remaining sequences base-pair relatively rapidly. The rate at which the probe “finds” the target, which is influenced by a number of factors (Table 2.10.2B), is therefore the limiting step in hybrid formation (Britten and Davidson, 1985). However, in practical terms hybridization rate is less important than hybrid stability, as in most protocols hybridization is allowed to proceed for so long that factors influencing rate become immaterial.

**Critical Parameters**

To be successful, a hybridization experiment must meet two criteria:

(1) **Sensitivity.** Sufficient probe DNA must anneal to the target to produce a detectable signal after autoradiography.

(2) **Specificity.** After the last wash, the probe must be attached only to the desired target sequence (or, with heterologous probing, family of sequences).

Parameters influencing these two criteria are considered in turn, followed by other miscellaneous factors that affect hybridization.

**Factors influencing sensitivity**

The sensitivity of hybridization analysis is determined by how many labeled probe molecules attach to the target DNA. The greater the number of labeled probe molecules that anneal, the greater the intensity of the hybridization signal seen after autoradiography.

**Probe specific activity.** Of the various factors that influence sensitivity, the one that most frequently causes problems is the specific ac-

**Preparation and Analysis of DNA****2.10.8**

Supplement 29

**EXHIBIT A**

e.g. expression vectors, the necessary common attribute is the ORF (SEQ ID NO: 2).

Weighing all factors including (1) that the full length ORF (SEQ ID NO: 2) is disclosed and (2) that any substantial variability within the genus arises due to addition of elements that are not part of the inventor's particular contribution, taken in view of the level of knowledge and skill in the art, one skilled in the art would recognize from the disclosure that the applicant was in possession of the genus of DNAs that comprise SEQ ID NO: 2.

**Conclusion:** The written description requirement is satisfied.

**Example 9: Hybridization**

**Specification:** The specification discloses a single cDNA ( SEQ ID NO:1) which encodes a protein that binds to a dopamine receptor and stimulates adenylate cyclase activity. The specification includes an example wherein the complement of SEQ ID NO: 1 was used under highly stringent hybridization conditions (6XSSC and 65 degrees Celsius) for the isolation of nucleic acids that encode proteins that bind to dopamine receptor and stimulate adenylate cyclase activity. The hybridizing nucleic acids were not sequenced. They were expressed and several were shown to encode proteins that bind to a dopamine receptor and stimulate adenylate cyclase activity. These sequences may or may not be the same as SEQ ID NO: 1.

**Claim:**

An isolated nucleic acid that specifically hybridizes under highly stringent conditions to the complement of the sequence set forth in SEQ ID NO: 1,

wherein said nucleic acid encodes a protein that binds to a dopamine receptor and stimulates adenylate cyclase activity.

**Analysis:**

A review of the full content of the specification indicates that the essential feature of the claimed invention is the isolated nucleic acid that hybridizes to SEQ ID NO: 1 under highly stringent conditions and encodes a protein with a specific function. The art indicates that hybridization techniques using a known DNA as a probe under highly stringent conditions were conventional in the art at the time of filing.

The claim is drawn to a genus of nucleic acids all of which must hybridize with SEQ ID NO: 1 and must encode a protein with a specific activity.

The search of the prior art indicates that SEQ ID NO: 1 is novel and unobvious.

There is a single species disclosed (a molecule consisting of SEQ ID NO: 1) that is within the scope of the claimed genus.

There is actual reduction to practice of the disclosed species.

Now turning to the genus analysis, a person of skill in the art would not expect substantial variation among species encompassed within the scope of the claims because the highly stringent hybridization conditions set forth in the claim yield structurally similar DNAs. Thus, a representative number of species is disclosed, since highly stringent hybridization conditions in combination with the coding function of DNA and the level of

skill and knowledge in the art are adequate to determine that applicant was in possession of the claimed invention.

**Conclusion:** The claimed invention is adequately described.

gi|2251107:452-1837 *Pyrococcus*  
SEQ ID NO:125  
10 20 30 40 50 60  
ATG---AAGAAGTTTGTG---CCCTGCTCATAACCATGTTTTCGTAGTGAGCATGG  
gtggtgcacatgaagtt-gaagtaccttgc-cttag---ttttgttggtgtg-gcttgcg

gi|2251107:452-1837 *Pyrococcus*  
SEQ ID NO:125  
70 80 90 100 110 120  
CT--GCCGTGACACAGC---CAGCTAGCGCCGCAAGTATTCGGAAGTCCGAAGAGGCGGC  
ataggectactctctgactccagtggtgtggtgccaagtactccgaactcgaagagggcggt

gi|2251107:452-1837 *Pyrococcus*  
SEQ ID NO:125  
130 140 150 160 170 180  
GTTATAATGCAGGCTTCTACTGGGACGTTCCAGCGGAGGAATCTGGTGGGATACATC  
gttataatgcaggccttctactgggatgttcccggagggggaatctggtgggacaccata

gi|2251107:452-1837 *Pyrococcus*  
SEQ ID NO:125  
190 200 210 220 230 240  
AGA-AGCAAGATACCGGAGTGGTACGAGGCTGGAATCTCCGCATCTGGATTCCGCCAGC  
agacagaaa-atcccgagtggtacgaagctggaatctcggcagataggattctctccagc

gi|2251107:452-1837 *Pyrococcus*  
SEQ ID NO:125  
250 260 270 280 290 300  
CAGCAAGGCGATGGGAGGAGCTTATTCAATGGGCTACGACCCATACGACTTCTTCGACCT  
tagcaaaagggatgggaggtgtgtatttcacgggctacgatccctacgatttctcttgacct

gi|2251107:452-1837 *Pyrococcus*  
SEQ ID NO:125  
310 320 330 340 350 360  
CGCCGAGTACAAACCAGAAGGGAACAGTTGAAACTCGCTTGGCTCAAAGCAGGAGCTTAT  
cggcgagtactatcagaagggaacagttggaagcgcttcgggtcaaaaggaggaaactggt

gi|2251107:452-1837 *Pyrococcus*  
SEQ ID NO:125  
370 380 390 400 410 420  
CAACATGATAAACACCGCCCATCGCTACGGCATAAAGGTCATAGCTGATATCGTCATAAA  
gaacatgataaacaccgcacactcctatggcataaagggtgatagcggacatagtcataaa

gi|2251107:452-1837 *Pyrococcus*  
SEQ ID NO:125  
430 440 450 460 470 480  
CCACCGCGCGGGCGGAGACCTCGAGTGGAAACCCGTTCTGGGGACTACACTGGACCGA  
ccaccgcgcggttgagaccttgagtggaaaccccttgtaaacactatacttggacaga

gi|2251107:452-1837 *Pyrococcus*  
SEQ ID NO:125  
490 500 510 520 530 540  
CTTCTCGAAGGTGGCTCGGGCAATATACCGCAACTACCTCGACTTCCACCCCAACGA  
cttctcgaaggtcgctcggtaataacaggccaactaccttgacttccacccaacga

gi|2251107:452-1837 *Pyrococcus*  
SEQ ID NO:125  
550 560 570 580 590 600  
GCTCAAGTGCTGTGACGAGGGGACATTTGAGGTTTCCAGACATAGCCACGAGAAGGA  
ggtcaagtgtgcatgagggacatttggtagcttccggacatcgccacagagaag-a

gi|2251107:452-1837 *Pyrococcus*  
SEQ ID NO:125  
610 620 630 640 650 660  
G-TGGGACGAGCACTGGCTCTGGGCGAGCCACGAGAGCTACGCCCGCTACTCTCAGGAGCA  
gctgggatcagtaactggtctctgggcaagcaatgagagctacgccgcatactctcggagca

gi|2251107:452-1837 *Pyrococcus*  
SEQ ID NO:125  
670 680 690 700 710 720  
TCGGCGTTGATGCCGTCGGCTTTGACTACGTAAGGGCTACCGAGCGTGGGTCTGTAAGG  
tagggatcgatgcattggcggtttcgactacgtcaaagggtacggagcggtgggtgttaag

gi|2251107:452-1837 *Pyrococcus*  
SEQ ID NO:125  
730 740 750 760 770 780  
ACTGGCTCAACTGGTGGGGCGGCTGGGCGCTCGGTGAGTACTGGACACGAACGTTGATG  
actggctcagctggtggggaggctgggcccgttggagagtactgggacacgaacggttgatg

gi|2251107:452-1837 *Pyrococcus*  
SEQ ID NO:125  
790 800 810 820 830 840  
CACTCCTCAATGGGCATACTCCAGCGGCGCCAAGGTCTTCGACTTCCCGCTCTACTACA  
cactcctcaactgggcatacagacagcgttgccaaggtctttgacttcccgcctctactaca

## EXHIBIT C

gi|2251107:452-1837 *Pyrococcus*  
SEQ ID NO:125

850 860 870 880 890 900  
.....|.....|.....|.....|.....|.....|  
AAATGGACGAGGCCTTTGACAAACACCAACATCCCGGCCTTGGTTTCATGCCCTCCAGAACG  
agatggacgaagcctttgacaacaccaacatccccgcttttggtttacgcctccagaacg

gi|2251107:452-1837 *Pyrococcus*  
SEQ ID NO:125

910 920 930 940 950 960  
.....|.....|.....|.....|.....|.....|  
GGGGAACCGTCGTCTCTCGCGACCCGTTCAAGGCCSTAACCTTGTAGCAAAACCACGACA  
gaggaacagtcgtttcccgcgatccccttcaaggcagtaacttttcgttgccaaccacgata

gi|2251107:452-1837 *Pyrococcus*  
SEQ ID NO:125

970 980 990 1000 1010 1020  
.....|.....|.....|.....|.....|.....|  
CCGATATAATCTGGAACAAGTACCCCTGCTTATGCTTTCATCCTCACCTACGAGGGCCAGC  
cagatataatctggaacaagtatcccgccttatggttccatccctacctaaggaggacagc

gi|2251107:452-1837 *Pyrococcus*  
SEQ ID NO:125

1030 1040 1050 1060 1070 1080  
.....|.....|.....|.....|.....|.....|  
CCGTCATATTTACCGCGACTACGAGGAGTGGCTCAACAAGGACAAGCTTAACAACCTAA  
ctgttatattttaccgcgactacgaggagtggttcaacaaggataagcttaacaacctta

gi|2251107:452-1837 *Pyrococcus*  
SEQ ID NO:125

1090 1100 1110 1120 1130 1140  
.....|.....|.....|.....|.....|.....|  
TCTGGATACACGACCCACCTCGCGGGTGAAGCACAGCATAGTCTACTACGACAGCGACG  
tctggatacacgagcaccttcgagggaagtacaaagatccctactactacgataacgatg

gi|2251107:452-1837 *Pyrococcus*  
SEQ ID NO:125

1150 1160 1170 1180 1190 1200  
.....|.....|.....|.....|.....|.....|  
AGCTCATCTTCTGAGGAACCGCCACTCCAAGACGCCGGCACTCATAACCTACATCAACC  
agctaatatttcattgaggggagggctacgggagcaagccgggcctcataactacataaacc

gi|2251107:452-1837 *Pyrococcus*  
SEQ ID NO:125

1210 1220 1230 1240 1250 1260  
.....|.....|.....|.....|.....|.....|  
TCGGCTCTAGCAAGGTCGAAAGTGGGTGTACGT--GC--CGAAGTTCCGCGGCCTGTC  
tcggaaacga-ctgggcccagagcgtgggtgaacgtcgggtcaagtttcgaggctacaca

gi|2251107:452-1837 *Pyrococcus*  
SEQ ID NO:125

1270 1280 1290 1300 1310 1320  
.....|.....|.....|.....|.....|.....|  
ATCCACGACTACACCGGCAACCTCGGAGGCTGGGTAGACAAGTACGTCCAGT-CGACCGG  
atccatgaatacacaggcaatctcggctggctgggttgacaggtgggttcagttacgat-gg

gi|2251107:452-1837 *Pyrococcus*  
SEQ ID NO:125

1330 1340 1350 1360 1370 1380  
.....|.....|.....|.....|.....|.....|  
CTGGGTCTATCTCGAAG-CTCCAGCTTACGAACCCGCCACCGGGGCATACGGGTACACCG  
atgggttaaat-gacggcacctctcatgatccagccaacggataattacggctactcag

gi|2251107:452-1837 *Pyrococcus*  
SEQ ID NO:125

1390 1400  
.....|.....|.....|.....|.....|  
TCTGGAGCTACTCCGGG-GTTGGATGA  
tctggagctac-gcaggcgtcggatga

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      10      20      30      40      50      60
SEQ ID NO:126
gi|2251108|dbj|BAA21130.1| alp  ---VHMKLKYLALVLL---AVASIGLL---STPVGA---AAKYSELEEGGVIMQA
MKRF-V-----A---LLITMFFVVSMAAVA---QPAS---AAKYSELEEGGVIMQA

      70      80      90     100     110     120
SEQ ID NO:126
gi|2251108|dbj|BAA21130.1| alp  FYWDVF---GGGIWWDTIQ---KIPewYD---AGISAIWIPPASKGMGG---YSMgyDPYDFFDLG
FYWDVPPAGG---IWWDTIR---SKIPEWY---EAGISAIWIPPASKGMGG---AYSMgyDPYDFFDLG

      130     140     150     160     170     180
SEQ ID NO:126
gi|2251108|dbj|BAA21130.1| alp  EYY---QKGTVETREFGSK---EELV---NMINTAHS---YGIKVIADIVINHRAGGDLEWNPFVNN---
EY---NQKGTVETREFGSKOE---L---INMINTAH---AYGIKVIADIVINHRAGGDLEWNPFV---GD

      190     200     210     220     230     240
SEQ ID NO:126
gi|2251108|dbj|BAA21130.1| alp  YTWTDfSKVASGKYTANYLDfHPNEVKCCDEGTFG---FPDIAHEKS---WDQY---WLWASN---E
YTWTDfSKVASGKYTANYLDfHPNEVKCCDEGTFG---GFPDIAHEK---EWDQ---HWLWAS---DE

      250     260     270     280     290     300
SEQ ID NO:126
gi|2251108|dbj|BAA21130.1| alp  SYAAYLRSIGI---DAWRFDYVKGYGAwVVN---DWT---SWWGGWAVGEYWDTNVDALLNWAYDS
SYAAYLRSIG---VDWRFDYVKGYGAwVV---KDWLN---WWGGWAVGEYWDTNVDALLNWAY---S

      310     320     330     340     350     360
SEQ ID NO:126
gi|2251108|dbj|BAA21130.1| alp  -GAKVFDFPLYyKMDEAFDNTNIPALVY---ALQNGGTVVSRDFFKAVTFVANHDTDIWNK
SGAKVFDFPLYyKMDEAFDNTNIPALV---DALQNGGTVVSRDFFKAVTFVANHDTDIWNK

      370     380     390     400     410     420
SEQ ID NO:126
gi|2251108|dbj|BAA21130.1| alp  YPAYAFILTYEGQPVIFYRDYEEWLNKDKLNNLIWIHE---HLAGGSTKIL---YYDN---DEL
YPAYAFILTYEGQPVIFYRDYEEWLNKDKLNNLIWIH---DHLAGGST---SIVYYD---SDEL

      430     440     450     460     470     480
SEQ ID NO:126
gi|2251108|dbj|BAA21130.1| alp  IFMREGY---G---SK---PGLITYINLNDWAE---RWVNVGS---KFAGYT---THEYTCN
IF-----VRNGDSKRPGGLITYINLNG---SSKVGRRV---YVPKFAG---ACIHEYTCN

      490     500     510     520     530
SEQ ID NO:126
gi|2251108|dbj|BAA21130.1| alp  LGGWVDRWVQYD-----GWVK---LTAP---PH---DPANGY---YGYs---VWSYA---GVG*
LGGWVD-----KYVESSGWV---YL---EAP---AYDPA---SGQYGY---TVWSY---CGVG-
```

## EXHIBIT D

Atty Docket No.: 56446-20061.00 /09010-108001/DIVER1530-5

## IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

Applicant : Walter Callen et al.                      Art Unit : 1652  
Serial No. : 10/081,872                              Examiner : Rebecca E. Prouty, Ph.D.  
Filed : February 21, 2002  
Title: Enzymes Having Alpha Amylase Activity and Methods of Use Thereof

MS RCE  
Commissioner for Patents  
P.O. Box 1450  
Alexandria, VA 22313-1450

DECLARATION UNDER 37 C.F.R. § 1.132

Sir:

1. I, Jay Short, am an expert in the field of molecular biology and enzyme development and was an expert at the time of the invention. I am presently employed as CEO and as research scientist at Diversa Corporation, San Diego, CA, assignee of the above-referenced patent application. My resume was provided with my previous declaration submitted in connection with this application.

2. It was considered routine by one skilled in the art to make multiple substitutions or multiple modifications in a nucleic acid sequence, such as a polypeptide coding sequence, and screen for variant enzymes, such as amylases, encoded by the modified nucleic acids. The state of the art at the time of the invention and the level of skill of the person of ordinary skill in the art for screening enzymes for amylase activity was very high. It would not have been necessary for the skilled artisan to understand which regions of the amylases encoded by nucleic acids used in the claimed methods could be modified to gain a function or activity, or, modified without loss of a function or activity, for example, amylase activity. It would not have been necessary for the skilled artisan to understand which specific regions or structural elements of the exemplary sequences were needed to have amylase function or activity (e.g., amylase activity) to routinely generate the genus of amylase-encoding nucleic acids used in the claimed methods. Methods for making and screening sequence modifications and enzyme fragments were sufficiently comprehensive, routine and predictable at the time of the invention to predictably generate a genus of amylase-encoding sequences without need of knowing which

sd-214331



Applicant : Walter Callen et al.  
Serial No. : 10/081,872  
Filed : February 21, 2002  
Page : 2 of 3

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108001/DIVER1530-5

specific regions or structural elements of a sequence or structure affected function or activity (e.g., amylase activity). Methods known at the time of the invention for modifying nucleic acid and polypeptide sequences in combination with high through-put enzyme (amylase) screening made methods that required previous knowledge of structural elements necessary for enzymatic activity obsolete and unnecessary. Methods known at the time of the invention for modifying nucleic acid and polypeptide sequences in combination with high through-put enzyme (amylase) screening known at the time of the invention, made methods that require previous knowledge of protein structure, including secondary or tertiary structure, active site sequences, and the like obsolete and unnecessary. At the time of the invention, high through-put *in vivo* (e.g., whole cell) nucleic acid expression and enzyme (amylase) screening protocols were well known in the art. The specification sets forth exemplary amylase screening assays to determine if a polypeptide is within the scope of the genus used in the claimed methods (see, e.g., Examples 1-2 and 5-10, of the specification). Using methods known in the art at the time of the invention it would not have been necessary to understand which specific regions of amylase structure needed to be modified to generate a genus of nucleic acids or polypeptides for practicing the claimed methods without undue experimentation. The specification presented to the skilled artisan a rational and predictable scheme for making the genus of amylases and amylase-encoding sequences, including a rational and predictable scheme for modifying the exemplary SEQ ID NO:125 with an expectation of obtaining an enzymatically active genus of amylase-encoding nucleic acids or amylase enzymes, including amylases. The specification provided sufficient guidance to one of ordinary skill in the art to make and use the genus of amylase-encoding nucleic acids to practice the methods of the invention.

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Applicant : Walter Callen et al.  
Serial No. : 10/081,872  
Filed : February 21, 2002  
Page : 3 of 3

Atty Docket No.: 56446-20061.00 /09010-  
108001/DIVER1530-5

I hereby declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code and that such willful false statements may jeopardize the validity of the application or any patent issued thereon.

Respectfully submitted

Date: \_\_\_\_\_

\_\_\_\_\_  
Jay Short

sd-214331

**CURRICULUM VITAE****NAME:** Jay M. Short, Ph.D.**EDUCATION:**

|             |  |
|-------------|--|
| 1981 - 1985 | Ph.D., Biochemistry<br>Case Western Reserve University, Cleveland, Ohio                    |
| 1980 - 1981 | Graduate Study, Macromolecular Science<br>Case Western Reserve University, Cleveland, Ohio |
| 1976 - 1980 | B.A. with Honors, Chemistry<br>Taylor University, Upland, Indiana                          |

**RESEARCH & PROFESSIONAL EXPERIENCE:**

|                       |  |
|-----------------------|--|
| <u>1999 - present</u> | <b>CEO, President, Chief Technology Officer &amp;<br/>Board of Directors</b><br>Diversa Corporation<br>San Diego, California           |
| <u>1998 - present</u> | <b>President, Chief Technology Officer &amp;<br/>Board of Directors</b><br>Diversa Corporation<br>San Diego, California                |
| 1997 - 1998           | <b>Executive Vice President, Chief Technology Officer &amp;<br/>Board of Directors</b><br>Diversa Corporation<br>San Diego, California |
| 1994 - 1997           | <b>Chief Technology Officer &amp;<br/>Board of Directors</b><br>Diversa Corporation<br>San Diego, California                           |
| <u>1990 - 1994</u>    | <b>President</b><br>Stratacyte, Inc.<br>La Jolla, California   |
| <u>1992 - 1994</u>    | <b>Vice President</b><br>R&D (Research) and Operations<br>Stratagene Cloning Systems<br>La Jolla, California                           |
| 1989 - 1992           | <b>Vice President</b><br>R&D (Research) and<br>Biological Operations<br>Stratagene Cloning Systems                                     |
| 1988 - 1989           | <b>Senior Staff Scientist</b><br>Research and Development<br>Stratagene Cloning Systems  |

**1985 - 1988**

**Staff Scientist**  
Research and Development  
Stratagene Cloning Systems

**1981 - 1985**

**Ph.D. Research**  
Case Western Reserve University  
Dr. Richard W. Hanson's Laboratory,  
Identification and characterization of the promoter for  
P-enolpyruvate carboxykinase.  
First Identification of a cAMP regulatory domain.

**1980 - 1981**

**Graduate Student Research**  
Case Western Reserve University  
Dr. Bruce Roe's Laboratory, Analysis of the cellulase activity of *Trichoderma viride*.

**TEACHING EXPERIENCE:**

Thesis Advisor (1988-1993), University of Uppsala, Sweden, Ph.D. for Michelle Alting-Mees  
Lecturer (1992), Committee for Advanced Scientific Education, Center for Drug  
Evaluation and Research, FDA.  
Faculty (1989), Transgenic Mouse Model and Its Application in Assessing  
*In Vivo* Mutagenesis, Genetic Toxicology Workshop (3rd Annual).  
Microbiological Associates Inc. Bethesda, MD.  
Faculty (1987), DNA Cloning and Expression. Physiology Society Workshop. San Diego, CA.  
Teaching Assist., (1981-1985). Molecular and Cellular Biology. Case Western  
Reserve University.  
Teaching Assist., (1981). Physiological Chemistry. Kent State Univ., Kent, OH.  
Teaching Assist., (1978-1980). Quantitative Analysis. Taylor University.

**AWARDS, PROFESSIONAL MEMBERSHIPS, ACCOMPLISHMENTS, AND ACTIVITIES:**

Visiting Scientist, International Centre of Insect Physiology and Ecology (ICIPE), Kenya (2002-2004)  
Science & Technology Committee, *BIOCOM San Diego*  
Advisory Board, IngleWood Ventures  
Finalists for UCSD Connect's Most Innovative New Product Award in the Biotechnology R&D Category  
Advisory Board, *Chemical & Engineering News*  
Board of Advisors and Founding Member, Division of Biological Sciences, *UCSD*  
Board Director, *BIOCOM San Diego*  
Chairman of the Board, Innovase  
Board Director, Zymetrics  
Board Director, Innovase  
Director at Large, *YPO (Young Presidents' Organization) San Diego*.  
2001 T-Sector Life Science Innovator Award.  
2001 Deloitte and Touche's Orange County / San Diego 2001 Technology "Fast 50".  
San Diego Entrepreneur of the Year 2001.  
YPO (Young Presidents' Organization) San Diego.  
YPO (Young Presidents' Organization) International.  
Finalist for San Diego Entrepreneur of the Year in 2000.  
Largest Biotechnology IPO raising over \$200MM.  
Founding management member of Diversa Corporation.  
Panel for Chemical Science & Technology for NIST, appointed by the National Research Council (1997-2000).  
Chairman (1993), Discussion Group, Society of Toxicology Conference.  
U.S. Committee Member for Evaluation of Biotechnology Research in Spain.  
Editor, Mutation Research.

UCSD Connect Program (1991) 1<sup>st</sup> Place Award for Innovation and Entrepreneurship in Biotechnology (over 50 competing biotech companies).  
 UCSD Connect Program (1990) 1<sup>st</sup> Place Award for Innovation and Entrepreneurship in Biotechnology.  
 Consultant for European Economic Community on Transgenic Toxicology Testing (91-94).  
 The New York Academy of Sciences.  
 Reviewer for *Proceedings of the National Academy of Sciences, Genetic Analysis Techniques, Analytical Biochemistry, & Nucleic Acids Research*.  
 American Association for the Advancement of Science.  
 American Chemical Society.  
 American Society of Biochemistry and Molecular Biology.  
 American Society of Microbiology.  
 Environmental Mutagenesis Society.  
 Society for Industrial Microbiology  
 Society of Toxicology.  
 Japanese Environmental Mutagen Society.  
 Who's Who Registry of Business Leaders (1994-1995)  
 American Men and Women of Science (1995)  
 NIEHS Peer Review Committee.  
 SBIR Study Section.  
 SBIR Annual Report (1993) Program Success Profile (Top 8 of 800 Companies).  
 Stratagene (1990) Innovation Award - Lambda ZAP<sup>®</sup> vector.  
 Stratagene (1990) Service Award  
 Stratagene (1991) Innovation Award - Big Blue<sup>®</sup> Transgenic Testing System.  
 Stratagene (1992) Most Innovative Award - Managers/Supervisors.  
 Institutional Animal Care and Use Committee (IACUC), Chairman and Institutional Official.  
 Award from the University of Victoria for Contributions to the Development of Short-term Transgenic Mutation Assays.  
 Nominated as Council Member for the U.S. Environmental Mutagen Society.  
 Board Director, *Stressgen (TSE), Victoria, BC, Canada*  
 Board Director & Treasurer, *Stressgen Therapeutics, Victoria, BC, Canada*  
 Board Director & Secretary, *Stressgen Therapeutics, Victoria, BC, Canada*  
 Board Director, *Diversa, La Jolla, CA*  
 Board Director, *Invitrogen, Carlsbad, CA*  
 Consultant, *Stratagene Cloning Systems, La Jolla, CA*  
 Consultant, *Micro Product Systems, Lynn, IN*  
 Reviewer for U.S. Congressional Office of Technology Assessment (OTA) on *The Human Genome Project and Patenting DNA Sequences*.

#### MEDIA:

ABC Discovery News, ABC San Diego Channel 10, BBC Radio, Bioinformed Newsletter, Biotechnology Newsletter, BioVentures View, Business Daily, Business Week, CEO Cast, Chemical Engineering, Chemical Week, Chemistry & Industry (UK), CNBC, CNN Science & Technology, dBusiness.com, Discovery Magazine, Forbes.com, Good Morning America, Horizon Air Magazine, Idea TV, Inside Business Radio Show, JAG Financial News, Los Angeles Times, NBC San Diego Channel 7/39, National Radio Report, New York Times, PiraInvestor.com, R&D Magazine, RTL German Television, Reuters, San Diego Business Transcript, San Diego Channel KUSI, San Diego Channel 10, San Diego Magazine, San Diego Union Tribune, Scientist, Time Magazine, Stewards' Watch, The Discovery Channel, The Motley Fool, Time Magazine, USA Today, Wall Street Journal, Wall Street Transcript, Washington Post

#### PATENTS:

DNA Cloning Vectors with *in vivo* Excisable Plasmids (1987).  
 Mutagenesis Testing Using Transgenic Animals Carrying Marker Genes (1987).  
 Mutagenesis Testing Using Transgenic Non-Human Animals Carrying Test

DNA Sequences (1987).  
Dietary and Hormonal Regulation of Expression of Exogenous Genes in Transgenic Animals Under Control of the Promoter of the Gene Phosphoenolpyruvate Carboxykinase (1988).  
A Transgenic Mouse for Measurement and Characterization of Mutation Induction *In Vivo* (1989).  
Rapid Screening Mutagenesis and Teratogenesis Assay (1989).  
A Combinatorial Approach to Regenerating the Immunoglobulin Repertoire in Prokaryotic Cells (1990).  
Transgenic Animal Models for *In Vivo* Mutagenesis Testing (1990).  
Polycos Vectors (1991).  
A Lambda Packaging Extract Lacking  $\beta$ -Galactosidase Activity (1991).  
A System for Regulation of Eukaryotic Genes (1991).  
Methods for Phenotype Creation from Multiple Gene Populations (1991).  
Transgenic Non-Human Animals Carrying Test DNA Sequences (1992).  
Mutagenesis Testing Using Transgenic Non-Human Animals Carrying Test DNA Sequences (1992).  
Selectable System Patent (1992).  
Polycos Mutagenesis Systems (1993).  
Use of Trans-acting Proteins for the Development of an *In Situ* Expression Screening System (1993).  
Enzyme Kits and Libraries (1995).  
Enzyme Activity Screening of Clones having DNA from Uncultivated Microorganisms (1995).  
Enzyme Tiered (1995).  
Method for Screening for Enzyme Activity (1995).  
Combined Enzyme Screening/Evolution (1995).  
Uncultured/Activity Screening (1995).  
Directed Evolution of Thermophilic Proteins (1995).  
Combinatorial Enzyme Development (Directed Mutagenesis) (1996).  
Protein Activity Screening of Clones having DNA from Uncultivated Microorganisms (1996).  
Production and Use of Normalized DNA Libraries (1996).  
Methods of DNA Shuffling with Polynucleotides Produced by Blocking or Interrupting a Synthesis or Amplification Process (1996).  
Method of Screening for Enzyme Activity (Biopanning) (1996).  
Directed Evolution of Thermophilic Enzymes (1996).  
Environmental Biopanning (1996).  
Combinatorial Enzyme Development (1996).  
Enzyme Activity Screening of Clones Having DNA from Uncultivated Microorganisms (1996).  
Normalized Samples/Libraries (1996).  
Reassembled Pools of Mutagenized DNA & Procedure (1996).  
Fluorescent-based Single Screening for Enzymes (1996).  
High Throughput Screening for Novel Enzymes (1997).  
Nucleotide Sequence of the *Aquifex aeolicus* Genome, Fragments Thereof, and Uses Thereof (1997).  
Screening for Novel Bioactivities (1997).  
Screening for Novel Compounds which Regulate Biological Interactions (1997).  
Method for Screening Enzyme Activity (1997).  
High Throughput Screening for Novel Enzymes (1997).  
"Discovery" (whole process, including uncultivated, normalized, biopanning, screening, evolving, etc.) (1997).  
Production of Enzymes Having Desired Activities By Mutagenesis (1999).  
Protein Activity Screening of Clones Having DNA from Uncultivated Microorganisms (1999).  
Method of DNA Reassembly by Interrupting Synthesis (1999).  
Production and Use of Normalized DNA Libraries (1999).  
Enzyme Kits and Libraries (1999).  
Screening for Novel Bioactivities (2000).  
Method for Screening for Enzyme Activity (2000).  
Screening for Novel Bioactivities (2000).  
Production and Use of Normalized DNA Libraries (2000).  
Method of Screening for Enzyme Activity (2000).  
Screening Methods for Enzymes and Enzyme Kits (2001).  
Saturation Mutagenesis in Directed Evolution (2001).  
High Throughput Screening for Novel Enzymes (2001).

Recombinant Bacterial Phytases and Uses Thereof (2001).  
 Methods Useful for Nucleic Acid Sequencing Using Modified Nucleotides Comprising Phenylboronic Acid (2001).  
 End Selection in Directed Evolution (2001)  
 Gene Expression Library Produced From DNA From Uncultivated Microorganisms and Method for Making the Same (2001)  
 Directed Evolution of Thermophilic Enzymes (2002)  
 Method for Screening for Enzyme Activity (2002)  
 Exonuclease-Mediated Gene Assembly in Directed Evolution (2002)  
 End Selection in Directed Evolution (2002)  
 Exonuclease-Mediated Gene Assembly in Directed Evolution (2002)  
 Screening for Novel Bioactivities (2002)  
 Method of DNA Shuffling with Polynucleotides Produced or Blocking or Interrupting Synthesis or Amplification Process (2002)  
 Production and Use of Normalized DNA Libraries (2002)  
 Sequence Based Screening (2002)  
 Non-Stochastic Generation of Genetic Vaccines (2002)  
 Over 100 Additional Pending Patent Applications Worldwide.

#### GRANTS AND CONTRACTS:

- \*Phase I Small Business Contract #N43-Am-62282. 1985 - 1986. P.I.  
Vectors and Techniques for Rapid DNA Sequencing.
- \*Phase II Small Business Contract #N43-Am-62282. 1988 - 1990. P.I.  
Vectors and Techniques for Rapid DNA Sequencing.
- \*Phase I Small Business Grant 2R43ES04484-02. 1986 - 1987. P.I.  
Identification of Genetic Lesions Leading to Mutations.
- \*Phase II Small Business Grant 2R43ES04484-02. 1989 - 1992. P.I.  
Identification of Genetic Lesions Leading to Mutations.
- \*1R01-ES04728-01A1. 1989 - 1992. (NIEHS) P.I.  
Animal Model for Identification of Genetic Lesions.
- \*Phase I Small Business Grant #R43GM42291-01. 1989. P.I.  
Switch Mechanism for Gene Expression in Transgenics.
- \*RFP NIH-ES-88-11. 1989-1994. (NIEHS) Co-I.  
Development of Mutagenesis Assays Using Transgenic Mice.
- \*Phase II Small Business Grant #2R44GM42291-02. 1990-1992. (DRG/NIH) P.I.  
Switch Mechanism for Gene Expression in Transgenics.
- \*Phase I Small Business Grant #1R43GM46585-01. 1991. (DRG/NIH) P.I.  
Generation of a Peptide Screening System Through the Development of  
Combinatorial-splicing "Polycos" Vectors.
- \*Phase I Small Business Grant #1R43CA57066-01. 1992. (NCI) P.I.  
Transgenic Rats: A Short-term Mutagenicity Assay for Multi-species Testing of Suspected Human Carcinogens.
- \*Phase I Small Business Grant #1R43GM48300-01. 1992. (DRG/NIH) P.I.  
Analysis of the Immunoglobulin Hypermutator Mechanism.
- \*Phase I Small Business Grant #1R43ES06146-01. 1992. (NIEHS) P.I.  
Selectable "Polycos" Shuttle Vectors for In Vivo Mutagenicity Testing.
- \*Phase II Small Business Grant #2R44GM46585-02. 1992-1994. (NIGMS) P.I.  
Peptide Screening Utilizing Combinatorial Polycos Vector.
- \*Phase I Small Business Grant #1R43RR08667-01. 1992-1993. (DRG/NIH) Co-I.  
A One-step PCR Cloning System Based on FLP Recombination.
- \*Phase II Small Business Grant #2R44CA57066-02. 1993-1995. (NCI) P.I.  
Transgenic Rats: Transgenic Rat Model for Mutagenicity Testing.
- \*Phase I Small Business Grant. 1993-1994. (NIH) Co-I.  
Transgenic Fish Model for Mutagenicity Testing.
- \*Phase II Small Business Grant (1994-1996). (NIH) P.I.  
"Polycos" Shuttle Vectors for Mutagenicity testing.
- \*Phase I Small Business Grant. 1994. (NIH) Co-I.  
Vector System for Studying Protein-Protein Interactions.

- \*CRADA with LLNL. 1994. (NIH) Co-I.  
Mouse and Rat Painting Probes.
- \*CRADA with FDA. 1994. (NIH) Co-I.  
Tamoxifen Testing in F-344 Rats.
- \*CRADA with NASA. 1994. (NIH) Co-I.  
Radiation Damage in the Microgravity Environment.

#### ABSTRACTS AND INVITED LECTURES:

Over 200 Abstracts and Invited Lectures.

#### PUBLICATIONS:

1. Yoo-Warren, H., Monahan, J.E., Short, J.M., Short, H., Bruzel, A., Wynshaw-Boris, A., Meisner, H.M., Samols, D., and Hanson, R.W. (1983) Isolation and Characterization of the Gene Coding for Cytosolic Phosphoenolpyruvate Carboxykinase (GTP) from the Rat. *Proc. Natl. Acad. Sci. U.S.A.*, 80:3656-3660.
2. Wynshaw-Boris, A., Lugo, T.G., Short, J.M., Fournier, R.E.K., and Hanson, R.W. (1984) Identification of cAMP Regulatory Region in the Gene for Rat Cytosolic Phosphoenolpyruvate Carboxykinase (GTP): Use of Chimeric Genes Transfected into Hepatoma Cells. *J. Biol. Chem.*, 259:12161-12169.
3. Wynshaw-Boris, A., Lugo, T.G., Short, J.M., Fournier, R.E.K., and Hanson, R.W. (1985) A Region of the Gene for Rat Cytosolic P-enolpyruvate Carboxykinase Confers cAMP Responsiveness to the HSV-thymidine Kinase Gene. In: *Membrane Receptors and Cellular Recognition*, (M. Czech and C.R. Kahn, eds.), Alan Liss Inc., New York, pp 339-346.
4. Wynshaw-Boris, A., Short, J.M., and Hanson, R.W. (1986) Characterization of the Phosphoenolpyruvate Carboxykinase (GTP) Promoter-Regulatory Region. I. Multiple Hormone Regulatory Elements and the Effects of Enhancers. *J. Biol. Chem.*, 261:9714-9720.
5. Short, J.M., Wynshaw-Boris, A., Short, H.P., and Hanson, R.W. (1986) Characterization of the Phosphoenolpyruvate Carboxykinase (GTP) Promoter-Regulatory Region. II. Identification of cAMP and Glucocorticoid Regulatory Domains. *J. Biol. Chem.*, 261:9721-9726.
6. Wynshaw-Boris, A., Short, J.M., and Hanson, R.W. (1986) The Determination of Sequence Requirements for Hormonal Regulation of Gene Expression. *Biotechniques*, 4:104-119.
7. Burns, D.M., Bhandari, G., Short, J.M., Sanders, P.G., Wilson, R.H., and Miller, R.E. (1986) Selection of a Rat Glutamine Synthetase cDNA Clone. *Biochemical and Biophysical Research Communications*, 134:146-151.
8. Hod, Y., Cook, J.S., Weldon, S.L., Short, J.M., Wynshaw-Boris, A., and Hanson, R.W. (1986) Differential Expression of the Genes for the Mitochondrial and Cytosolic Forms of P-enolpyruvate Carboxykinase Gene. In: *Metabolic Regulation: Application of Recombinant DNA Techniques*, (A.G., Goodridge and R.W. Hanson eds.), Annals of the New York Academy of Sciences, New York, Vol. 278, pp. 31-45.
9. Wynshaw-Boris, A., Short, J.M., and Hanson, R.W. (1987) *cls* - acting Regulatory Elements in Hormonally Responsive Genes. In: *Progress in Nucleic Acid Research and Molecular Biology* (W.E. Cohn and K. Moldave eds.), Academic Press, Inc., Orlando, Florida, 34:59-87.
10. Bullock, W., Fernandez, J.M., and Short, J.M. (1987) XL1-Blue: A High Efficiency Plasmid Transforming *recA E.coli* Strain With  $\beta$ -Galactosidase Selection. *Biotechniques*, 5:60-64.
11. Short, J.M., Fernandez, J.F., Sorge, J.A., and Huse, W. (1988) Lambda ZAP<sup>®</sup>: A Bacteriophage Lambda Expression Vector With *In Vivo* Excision Properties. *Nucleic Acids Res.*, 16:7583-7600.



12. Short, J.M. (1988) Book Review: Vectors - A Survey of Molecular Cloning Vectors and Their Uses. Raymond L. Rodriguez and David T. Denhardt, eds, Butterworths, Stoneham, MA. *Genomics*, 2:270-271.
13. Short, J.M., and Pollard, A. (1988) Gigapack XL: Size Selective Packaging Extract. *Strategies in Mol. Biol.*, 1:5-7.
14. Kretz, P.L., and Short, J.M. (1989) Gigapack II: A Restriction Deficient (*mcrA*-, *B*-, *hsd*-, *mrr*-) Lambda Packaging Extract. *Strategies in Mol. Biol.*, 2(2):25-26.
15. Kretz, P.L., Reid, C.H., Greener, A., and Short, J.M. (1989) Effect of Lambda Packaging Extract *Mcr* Restriction Activity on DNA Cloning. *Nucleic Acids Res.* 17:5409.
16. Sastry, L., Altling-Mees, M., Huse, W.D., Short, J.M., Sorge, J.A., Hay, B.N., Janda, K.D., Benkovic, S.J., and Lerner, R.A. (1989) Cloning of the Immunological Repertoire in *E. coli* for Generation of Monoclonal Catalytic Antibodies I. Construction of a  $V_H$  Specific cDNA Library. *Proc. Natl. Acad. Sci. U.S.A.*, 86:5728-5732.
17. Short, J.M. (1989) The Use of Lambda Phage Shuttle Vectors in Transgenic Mice for Development of a Short Term Mutagenicity Assay. In: *Fifth International Conference on Environmental Mutagens*, Alan Liss, Inc., New York, Part A:335-367. Article and Lecture.
18. Altling-Mees, M., and Short, J.M. (1989) pBluescript II: Gene Mapping Vectors. *Nucleic Acids Res.*, 17:9494.
19. Shopes, B., Altling-Mees, M., Amber, J.R., Ardourel, D., Callahan, M., Detrick, J., Hay, B.N., Hogrefe, H.H., Greener, A., Gross, E.A., Kubitz, M.M., Mullinax, R.L., Wilson, C., Short, J.M., and Sorge, J.A. (1990) Bacteriophage Immuno-expression Libraries: An Emerging Technology for the Identification and Production of Monoclonal Antibodies. *Antibody Engineering, New Tech. & Application Implications*. pp. 98-101.
20. Altling-Mees, M., Amberg, J., Ardourel, D., Elgin, E., Greener, A., Gross, E.A., Kubitz, M., Mullinax, R.L., Short, J.M., and Sorge, J.A. (1990) Monoclonal Antibody Expression Libraries: A Rapid Alternative to Hybridomas. *Strategies in Mol. Biol.*, 3:1-9.
21. Kohler, S., Provost, S., Dyaico, M., Sorge, J., and Short, J.M. (1990) Development of a Short-term, *In Vivo* Mutagenesis Assay: The Effects of Methylation on the Recovery of a Lambda Phage Shuttle Vector from Transgenic Mice. *Nucleic Acids Res.*, 18:3007-3013.
22. Kohler, S., Provost, G.S., Kretz, P.L., Fieck, A., and Short, J.M. (1990) An *In Vivo* Assay Using Transgenic Mice to Analyze Spontaneous and Induced Mutations at the Nucleic Acid Level. *Strategies in Mol. Biol.*, 3:19-21.
23. Kretz, P., Kohler, S., and Short, J.M. (1990) The Effect of *E. coli* Minute 98 Locus on DNA Containing Eukaryotic Modifications. *Strategies in Mol. Biol.*, 3:21-22.
24. Mullinax, R.L., Gross, E.A., Amberg, J., Hogrefe, H., Kubitz, M., Greener, A., Altling-Mees, M., Ardourel, D., Hay, B.N., Short, J.M., Sorge, J.A., and Shopes, B. (1990) Identification of Human Antibody Fragment Clones Specific for Tetanus Toxin in a Bacteriophage Lambda Immuno-Expression Library. *Proc. Natl. Acad. Sci. U.S.A.*, 87:8095-8099.
25. Cline, J., Lundberg, K., Nielson, K., Sorge, A., Short, J.M., and Mathur, E.J. (1990) StrataClean Resin: Non-Toxic Protein Extraction. *Strategies in Mol. Biol.*, 4(4):49-51.
26. Mullinax, R.L., Gross, E.A., Amber, J.R., Hay, B.N., Hogrefe, H.H., Kubitz, M.M., Greener, A., Altling-Mees, M., Ardourel, D., Short, J.M., Sorge, J.A., and Shopes, B. (1990) Human Antibody Clones Isolated From a Bacteriophage Lambda Immunoexpression Library. *Strategies in Mol. Biol.*, 4(4):51-52.
27. Provost, G.S., Kohler, S.W., Fieck, A., Kretz, P.L., Molina, T., and Short, J.M. (1990) Short-term Germ Line and Somatic Cell Mutagenesis Testing With *LacI* Lambda Phage Shuttle Vectors in Transgenic Mice. *Strategies in Mol. Biol.*, 4(4):55-56.

28. Kohler, S.W., Provost, G.S., Kretz, P.L., Fieck, A., Sorge, J.A., and Short, J.M. (1990) The Use of Transgenic Mice for Short Term, *In Vivo* Mutagenicity Testing. *Genetic Analysis Techniques*, 7(8):212-218.
29. Shopes, B., Mullinax, R.L., Amber, J.R., Gross, E.A., Hay, B.N., Hogrefe, H.H., Kubitz, M.M., Greener, A., Alting-Mees, M., Ardourel, D., Short, J.M., and Sorge, J.A. (1990) ImmunoZAP<sup>®</sup> Bacteriophage Libraries: A New Technology for the Identification and Expression of Monoclonal Antibodies. *Biotech USA Conference Proceedings*, pp.332-341.
30. Raleigh, E.A., Benner, J., Bloom, F., Braymer, H.D., DeCruz, E., Dharmalingam, K., Heitman, J., Noyer-Weldner, M., Piekarowicz, A., Kretz, P.L., Short, J.M., and Woodcock, D. (1991) Nomenclature Relating to Restriction of Modified DNA in *Escherichia coli*. *Journal of Bacteriology*, 173(8):2707-2709.
31. Kretz, P., Kohler, S., and Short, J.M. (1991) Identification and Characterization of a Gene Responsible for Inhibiting Propagation of Methylated DNA Sequences in *mcrA*, *mcrB1* *E. coli* Strains. *Journal of Bacteriology*, 173:4707-4716.
32. Kohler, S.W., Provost, G.S., Fieck, A., Kretz, P.L., Bullock, B., Sorge, J.A., Putman, D., and Short, J.M. (1991) Spectra of Spontaneous and Induced Mutations Using a Lambda ZAP<sup>®</sup> LacI Shuttle Vector in Transgenic Mice. *Proc. Natl. Acad. Sci. U.S.A.*, 88(18):7958-7962.
33. Wyborski, D., and Short, J.M. (1991) Analysis of Inducers of the *E.coli* Lac Repressor System in Mammalian Cells and Whole Animals. *Nucleic Acids Research*, 19:4647-4653.
34. Lundberg, K.L., Shoemaker, D.D., Adams, M.W.W., Short, J.M., Sorge, J.A., and Mathur, E.J. (1991) High Fidelity Amplification With a Thermostable DNA Polymerase Isolated from *Pyrococcus Furiosus*. *Gene*, 108:1-6.
35. Kohler, S.W., Provost, G.S., Fieck, A., Kretz, P.L., Bullock, W.O., Putman, D.L., Sorge, J.A., and Short, J.M. (1992) Analysis of Spontaneous and Induced Mutations in Transgenic Mice Using a Lambda ZAP<sup>®</sup>/LacI Shuttle Vector. *Environmental and Molecular Mutagenesis*, 18:316-321.
36. Fieck, A., Wyborski, D., and Short, J.M. (1992) Modifications of the *E. coli* Lac Repressor for Expression in Eukaryotic Cells: Effects of Nuclear Signal Sequences on Protein Activity and Nuclear Accumulation. *Nucleic Acids Research*, 20:1785-1791.
37. Hay, B., and Short, J.M. (1992) ExAssist<sup>TM</sup> Helper Phage and SOLR<sup>TM</sup> Cells for Lambda ZAP<sup>®</sup> II Excisions. *Strategies in Mol. Biol.*, 5:16-18.
38. Short, J.M. (1992) Tissue Specific Mutagenesis in Transgenic Mice. *The Toxicology Forum, 1992 Annual Winter Meeting*, pp.79-109.
39. Alting-Mees, M.A., Sorge, J.A., and Short, J.M. (1992) pBluescript II: Multifunctional Cloning and Mapping Vectors. *Methods in Enzymology*, 216:483-495.
40. Short, J.M., and Sorge, J.A. (1992) *In Vivo* Excision Properties of Bacteriophage Lambda ZAP<sup>®</sup> Expression Vectors. *Methods in Enzymology*, 216:495-508.
41. Short, J.M. (1992) Transgenic Animals for Carcinogenicity and Genotoxicity Testing. *Biotechnology International, The Global Review of Industry Manufacture and Application 1992*. Section.2. pp. 91-99.
42. Provost, G.S., Hamner, R., Kretz, P.L., and Short, J.M. (1992) Response to the Commentary Article: Comparison of Mutation Frequencies Obtained Using Transgenes and Specific-locus Mutation Systems in Male Mouse Germ Cells. *Mutation Research*, 298:145-147.
43. DuCoeur, L.C., Wyborski, D.L., and Short, J.M. (1992) Control of Gene Expression in Eukaryotic Cells Using the Lac Repressor System. *Strategies in Mol. Biol.*, 5(3):70-72.
44. Jerpseth, B., Greener, A., Short, J.M., Viola, J., and Kretz, P.L. (1992) XL1-Blue MRF: *mcrA*-, *mcrCB*-, *mrr*-, *HsdRMS*- derivative of XL1-Blue. *Strategies in Mol. Biol.*, 5(3):81-83.

45. Altling-Mees, M., Hoener, P., Ardourel, D., Sorge, J., and Short, J.M. (1992) ZAP Express™ and pBK-CMV, pBK-RSV Phagemid Vectors for Prokaryotic and Eukaryotic Expression. *Strategies in Mol. Biol.*, 5(3):58-61.
46. Short, J.M., Provost, G.S., Kretz, P.L., and Dyaico, M.J. (1992) Overview of the Big Blue® *In Vivo* Mutagenesis Assay. *Mammalian Mutagenesis Study Group Communication -- JEMS.MMS*, 6:73-89.
47. Lundberg, K.S., Kretz, P.L., Provost, G.S., and Short, J.M. (1993) The Use of Selection in Recovery of Transgenic Targets for Mutation Analysis. *Mutation Research Letters*, 301/2:99-105.
48. Mirsalis, J., Provost, G.S., Matthews, C., Hamner, R., Schindler, J.E., O'Loughlin, K., MacGregor, J.T., and Short, J.M. (1993) Induction of Hepatic Mutations in *LacI* Transgenic Mice. *Mutagenesis*, 8:265-271.
49. Altling-Mees, M.A., Vaillancourt, P., and Short, J.M. (1993) Phagemids and Other Hybrid Vectors. In: *Plasmids: A Practical Approach*. (ed. K. Hardy). IRL Press, pp. 197-223.
50. Provost, G.S., Kretz, P.L., Dyaico, M., Lundberg, K., and Short, J.M. (1993) Transgenic Systems for *In Vivo* Mutation Analysis. *Mutation Research*, 288:133-149.
51. Hedden, V., Callen W., Short, J.M., and Kretz, K. (1993) Improved Sequence Analysis of Mutations Identified With the Big Blue® System. *Strategies in Mol. Biol.*, 6:27-28.
52. Jerpseth, B., Greener, A., Short, J.M., Viola, J., and Kretz, P.L. (1993) New Restriction-Minus Derivatives of XL1-Blue *E. coli* Cells. *Strategies in Mol. Biol.*, 6:24.
53. Vaillancourt, P., Wyborski, D.L., and Short, J.M. (1993) The FLASH® CAT Kit: A Fast, Sensitive CAT Assay Without Radioactivity. *Strategies in Mol. Biol.*, 5:17-19.
54. Short, J.M., Dyaico, M.J., Provost, G.S., Kretz, P.L., Rogers, B.J., Ardourel, D., Wyborski, D.L. and Moores, J.C. (1993) Transgenic Mice and Rats for Tissue Specific Mutation Analysis. *JEMS*, 22:45-46.
55. Altling-Mees, M.A., and Short, J.M. (1993) Polycos Vectors: Filamentous Phage Packaging Using Lambda Extracts. *Gene*, 137:93-100.
56. Piegorsch, W.W., Lockhart, A.C., Margolin, B.H., Tindall, K.R., Gorelick, N.J., Short, J.M., Carr, G.J., and Shelby, M.D. (1994) Sources of Variability in Data from a *lacI* Transgenic Mouse Mutation Assay. *Environmental & Molecular Mutagenesis*, 23:17-31.
57. Dyaico, M., Provost, G.S., Kretz, P.L., Ransom, S.L., Moores, J.C. and Short, J.M. (1994) The Use of Shuttle Vectors for Mutation Analysis in Transgenic Mice and Rats. *Mutation Research*, 307:461-478.
58. Wyborski, D.L., Malkhosyan, S., Moores, J.C., Dyaico, M.J., and Short, J.M. (1994) Rat2 Cell Line for *In Vitro* Mutagenicity Testing. *Strategies in Mol. Biol.*, 7(2):55-56.
59. Provost, G.S. and Short, J.M. (1994) Characterization of Mutations Induced by Ethylnitrosourea in Seminiferous Tubule Germ Cells of Transgenic B6C3F1 Mice. *Proc. Natl. Acad. Sci. U.S.A.*, 91:6564-6568.
60. Kretz, P.L., Wells, S., and Short, J.M. (1994) Gigapack III: A Single Tube *In Vitro* Lambda Packaging Extract. *Strategies in Molecular Biology* 7:44-45.
61. Knoll, A., Jacobson, D., Kretz, P., Lundberg, K., Short, J., and Sommer, S. (1994) Tissue-Specific Patterns of Spontaneous *LacI* Mutations Recovered From Transgenic Mice. *Mutation Research* 311:57-67.
62. Ashby, J., Short, J.M., Jones, N.J., Lefevre, P.A., Martin, E., Parry, J.M., Burnette, K., Glickman, B.W., and Tinwell, H. (1994) Mutagenicity of o-anisidine to the bladder of *lacI*-transgenic B6C3F1 mice: Absence of <sup>14</sup>C or <sup>32</sup>P bladder DNA adduction. *Carcinogenesis* 15:2291-2296.
63. Altling-Mees, M. and Short, J.M. (1994) Rapid Excision Systems. *Strategies in Molecular Biology* 7(3):70-72.

64. Snead, M., Kretz, P.L., Short, J.M. (1994) Methods for Generating Plant Genomic Libraries. *Plant Molecular Biology Manual* H1:1-19.
65. Rogers, B., Provost, G.S., Young, R., Putman, D.L., and Short, J.M. (1995) Intralaboratory Optimization and Standardization of Mutant Screening Conditions Used for a Lambda/*LacI* Transgenic Mouse Mutagenesis Assay (I). *Mutation Research* 327:57-66.
66. Young, B., Rogers, B., Provost, G.S., Short, J.M., and Putman, D. (1995) Interlaboratory Comparison of Liver Spontaneous Mutant Frequency from Lambda/*LacI* Transgenic Mice (Big Blue®) (II). *Mutation Research* 327:67-73.
67. Callahan, J. and Short, J.M. (1995) Transgenic  $\lambda$ /*lacI* Mutagenicity Assay: Statistical Determination of Sample Size. *Mutation Research* 327:201-208.
68. Wyborski, D.L., Malkhosyan, S., Moores, J.C., Dyaico, M.J., Perucho, M. and Short, J.M. (1995) Development of a Rat Cell Line Containing a Lambda Shuttle Vector for *In Vitro* Mutagenicity Testing. *Mutation Research* 334:161-165.
69. Snead, M.A., Kretz, P.L., and Short, J.M. (1995) Methods for Generating Plant Genomic Libraries. *Plant Molecular Biology Manual*, Kluwer Academic Publishers: Belgium (H1, 1-19).
70. Robertson, D.E., Mathur, E.J., Swanson, R.V., Marrs, B.L., and Short, J.M. (1996) The Discovery of New Biocatalysts From Microbial Diversity. *SIM News* 46:3-8.
71. Knoll, A., Jacobson, D.P., Nishino, H., Kretz, P.L., Short, J.M., and Sommer, S. (1996) A Selectable System for Mutation Detection in the Big Blue® *lacI* Transgenic Mouse System: What Happens to the Mutational Spectrum Over Time. *Mutation Research* 352:9-22.
72. Wyborski, D.L., DuCoeur, L.C., and Short, J.M. (1996) The Effect of Chromosome Position and Operator Placement on Lac Repressor Control in Eukaryotic Cells and Transgenic Mice. *Environmental and Molecular Mutagenesis*. 28:447-458.
73. Sick, A.J., Fernandez J. and Short, J.M. (1996) Multiple Purpose Cloning Vectors. *Molecular Biology* (published).
74. Snead, M., Altling-Mees, M.A., and Short, J.M. (1997) cDNA Library Construction for the Lambda ZAP® - Based Vectors. *Methods in Molecular Biology, cDNA Library Protocols*. Humana Press 69:39-51.
75. Snead, M., Altling-Mees, M.A., and Short, J.M. (1997) Clone Excision Methods for the Lambda ZAP® - Based Vectors. *Methods in Molecular Biology, cDNA Library Protocols*. Humana Press 69:53-60.
76. Short, J.M. (1997) Recombinant Approaches for Accessing Biodiversity. *Nature Biotechnology* 15:1322-1323.
77. Nichols, W.S., Geller, S.A., Edmond, V.J., Dyaico, M.J., Sorge, J.A., and Short, J.M. (1998) Hepatocarcinogenesis (Z#2) / mutagenesis during initiation stage. *Mutation Research* 398:143-149.
78. Snead, M., Altling-Mees, M.A., and Short, J.M. (1998) cDNA Library Construction for Lambda ZAP® - Based Vectors. *Methods in Molecular Biology, Plant Virology Protocols*. Humana Press 81:255-267.
79. Bruggeman, T., Short, J.M., and Simms, P. (1998) Diversa: Catalyzing a Revolution. *Industrial Biotech News* 1(1):4,14-15.
80. Deckert, G., Warren, P.V., Gaasterland, T., Young, W.G., Lenox, A.L., Graham, D.E., Overbeek, R., Snead, M., Keller, M., Aujay, M., Huber, R., Feldman, R.A., Short, J.M., Olsen, G.J., and Swanson, R.V. (1998) The Complete Genome of the Hyperthermophilic Bacterium *Aquifex aeolicus*. *Nature* 392:353-358.
81. Li, J., Robertson, D.E., Short, J.M., Wang, P.G. (1999) Chemical and enzymatic synthesis of glycoconjugates. 5: One-pot regioselective synthesis of bioactive galactobiosides using a CLONEZYME thermophilic glycosidase library. *Bioorganic & Medicinal Chemistry Letters* Jan 4; 9 (1):35-8

82. Snead, M.A., Alting-Mees, M.A., Short, J.M. (2000) cDNA Library Construction for the Lambda ZAP® - Based Vectors. *Nucleic Acid Protocols Handbook, Humana Press Part V*:355-365.
83. Sehgal, A.C., Callen W., Mathur E. J., Short, J.M., Kelly, R.M. (2001) Carboxylesterase from *Sulfolobus Solfataricus* P1. *Methods in Enzymology* 330:461-471.
84. Cady, S.G., Bauer, M.W., Callen, W., Snead, M.A., Mathur, E.J., Short, J.M., Kelly, R.M. (2001) Beta-Endoglucanase from *Pyrococcus Furiosus*. *Methods in Enzymology* 330:346-354.
85. Miller, E.S., Kimberley, Parker, N., Liebl, W., Lam, D., Callen, W., Snead, M.A., Mathur, E.J., Short, J.M., Kelly, R.M. (2001) Alpha-D-galactosidases from *Thermotoga* Species. *Methods in Enzymology* 330:246-260.
86. Chhabra, S., Parker, K.N., Lam, D., Callen, W., Snead, M.A., Mathur, E.J., Short, J.M., Kelly, R.M. (2001) Beta-mannanases from *Thermotoga* Species. *Methods in Enzymology* 330:224-238.
87. Parker, K.N., Chhabra, S.R., Lam, D., Callen, W., Duffaud, G.D., Snead, M.A., Short, J.M., Mathur, E.J., Kelly, R.M. (2001) Galactomannanases Man2 and Man5 from *Thermotoga* species: growth physiology on galactomannans, gene sequence analysis, and biochemical properties of recombinant enzymes. *Biotechnology and Bioengineering* Nov 5;75 (3):322-33
88. Gray, K.A., Richardson, T.H., Kretz, K., Short, J.M., Bartnek, F., Knowles, R., Kan, L., Swanson, P.E., Robertson, D.E. (2001) Rapid Evolution of Reversible Denaturation and Elevated Melting Temperature in a Microbial Haloalkane Dehalogenase. *Advanced Synthesis & Catalysis* 2001, 343:607-617.
89. Richardson, T.H., Tan, X., Frey, G., Callen, W., Cabell, M., Lam, D., Macomber, J., Short, J.M., Robertson, D., Miller, C. (2002) A Novel, High Performance Enzyme for Starch Liquefaction: Discovery and Optimization of a Low pH, Thermostable  $\alpha$ -amylase. *Journal of Biological Chemistry* 2002, 277(29), 26501-26507.
90. Zengler, K., Toledo, G., Rappe, M., Elkins, J., Mathur, E.J., Short, J.M., Keller, M. (2002) Cultivating the Uncultured. *Proceedings of the National Academy of Sciences of the United States of America* (2002), 99(24), 15681-15686.
91. Murphy, K.M., Broman, K.W., Ziegler, J.S., Wyborski, D.L., Joe, L.K., Smith, D.W., Thurston, L.M., Stevenson, S.E., McClelland, M., Short, J.M., Mathur, E.J., Varley, J.D. (2002) Successful Breeding Among Free-Ranging 10-Month-Old Gray Wolves (*Canis Lupus*) in Yellowstone National Park, Wyoming. *Yellowstone Center for Resources* (2002). Kerry M. Murphy ([kerry\\_murphy@nps.gov](mailto:kerry_murphy@nps.gov)), P.O. Box 168, Yellowstone National Park, WY 82190.